How to clone Targets into pCP5b for Yeast TALEN Assay:

1. Oligos

Order the Sense and Anti-sense oligos from IDT, usually I order 25nmole with standard desalting

For Example:

On sense Oligo: GATC is the BglII compatible overhang

t is the t at the -1 position for TALEN 1 AAAAAAAAAAAAAAAAAA is TALEN1

CTCTCTCTCTCTCT is the spacer

a is complementary to the t at -1 position for TALEN2

On Anti-sense Oligo: CTAG is the SpeI compatible overhang

t is the t at the -1 position for TALEN2 CCCCCCCCCCCCCCCCCCC is TALEN2 GAGAGAGAGAGAGAG is the spacer

TTTTTTTTTTTTTTTTTTT is complementary to TALEN1 a is complementary to the t at -1 position for TALEN1

2. Vector

Digest pCP5b with BglII+SpeI+CIP
10ug pCP5b
30µl 10X Buffer 2 (NEB)
3µl 100X BSA (NEB)
10µl BglII (NEB)
10µl SpeI (NEB)
10µl CIP (NEB)
? µl water (up to 300µl)

Incubate at 37°C for 1h

Expected Sizes: 10370bp and 2493bp Gel purify the 10kb band (use Qiagen kit) Quantify recovered DNA

3. Phosphorylation and annealing

Suspend oligos to $100\mu \text{M}$ in TE

Mix:

3μl Sense oligo

3μl Anti-sense oligo

3μl 10X T4 DNA Ligase Buffer (NEB)

2μl T4 Polynucleotide Kinase (NEB)

19µl water

Incubate at 37°C for 30min

Add 4µl 0.5M NaCl

Boil for 2min

Cool slowly on bench to room temperature

Dilute 1µl into 499µl water

4. Ligation

Mix:

l μl diluted annealed oligos 40ng pCP5b/BglII+SpeI+CIP 2μl 10X Ligase Buffer (NEB) 1μl T4 DNA Ligase (NEB) ? μl water (up to 20 μl)

Incubate at Room temperature 2h Transform $5\mu l$ into DH5 α Plate on LB+Ap

Sequence clones with P789: ACAGAAAAGCAGGCTGGGAAGCA